## The Antibacterial Activity of *Angelica glauca* Edgew (Apiaceae) in The Form of Silver Nanoparticles Zahraa Sedeeq Qasim<sup>\*,1</sup> and Khadeeja Younis Abid<sup>2</sup>

<sup>1</sup>Department of Clinical Laboratory Science, College of Pharmacy, University of Mosul, Mosul, Iraq. <sup>2</sup>Department of Pharmacognosy, College of Pharmacy, University of Mosul, Mosul, Iraq.

## Abstract

Angelica glauca (Apiaceae) is traditionally an over-the-counter herbal preparation; although nowadays, nanotechnology formulates silver nanoparticles (AgNPs) from Angelica glauca. Utilization of green chemistry through nanoparticle formulations contributed to pharmaceutical properties enhancements in plants' crude extracts. The present study aims to evaluate the antibacterial activity of the synthesized silver nanoparticles (AgNPs) from Angelica glauca crude methanolic extract and perform phytochemicals assay screening Angelica glauca AgNPs by scan electron microscope (SEM), and finally *in vitro* antibacterial assay of the methanolic extract and silver nanoparticles. After soxhlet extraction, the recovered crude methanolic extract of dried Angelica glauca roots was subjected to nanoparticles derivatization, additionally, the AgNPs were analyzed with SEM. Five metabolites were detected, the *in vitro* antibacterial assay was performed using disc diffusion method against four selected microbial species. The extract yield was 13.3g, and the phytochemical analysis revealed the presence of alkaloids, flavonoids, tannins, steroids, and coumarins. The detected particle size was 70.90 nm  $\pm$  13.60 (mean  $\pm$  SD). The crude extract's antibacterial potential was enhanced by the nanoparticulation. The results of this study display the AgNPs semisynthetic product as an approach to improve the crude extracts' bioactivities. In conclusion, Angelica glauca contains several bioactive phytochemicals which can be used in curing health-related problems.

Keywords: Antimicrobial activity, Angelica glauca, Methanolic extract, Silver nanoparticles.

## النشاط المضاد للبكتريا لحشيشة الملاك ايدجو ( الفصيلة الخيمية) في شكل حبيبات فضة نانوية زهراء صديق قاسم \*٬ ه خديجة بونس عدرً

زهراء صديق قاسم \*<sup>، ا</sup>و خديجة يونس عبد <sup>٢</sup> فرع العلوم المختبرية السريرية ، كلية الصيدلة ، جامعة الموصل ، موصل، العراق فرع العقاقير ، كلية الصيدلة ، جامعة الموصل ، موصل، العراق

## الخلاصة

حشيشة الملاك (الفصيلة الخيمية) مستحضر طبي عشبي تقليدي بدون وصفة طبية، اما في الوقت الحاضر فان تقنية النانو صنعت حبيبات نانوية فضية من حشيشة الملاك باستخدام طريقة التخليق الأخضر من خلال تخليق الحبيبات النانوية والتي يعزى لها زيادة الصفة الصيدلانية في المستخلصات الخام النباتية. هدفت هذه الدراسة لتقبيم النشاط المضاد للبكتريا لحبيبات الفضة النانوية المصنعة من المستخلص الخام الميثانولي لحشيشة الملاك لتخليق حبيبات الفضة النانوية لحشيشة الملاك من مستخلصها الميثانولي ، كشف المجاميع الكيميائية النباتية ،التحري عن الحبيبات الفضة النانوية بالميكروسكوب الإلكتروني, أخيرًا تقبيم النشاط المضاد للبكتريا خارج الجسم الحي لحشيشة لملاك لاشكال المستخلص الميثانولي و الفضة النانوية بالميكروسكوب الإلكتروني, أخيرًا تقبيم النشاط المضاد للبكتريا خارج الجسم الحي لحشيشة لملاك لإشكال المستخلص الميثانولي و حبيبات الفضة النانوية بالميكروسكوب الإلكتروني, أخيرًا تقبيم النشاط المضاد للبكتريا خارج الجسم الحي لحشيشة لملاك لإشكال المستخلص الميثانولي و حبيبات الفضة النانوية بالميكروسكوب الإلكتروني, أخيرًا تقبيم النشاط المضاد للبكتريا خارج الجسم الحي لحشيشة لملاك الميثانولي المناء الميثانولي و حبيبات الفضة النانوية بالميكروسكوب والكتروني أخيرًا تقبيم النشاط المضاد للبكتريا خارج الجسم الحي لحيثيشة الملاك الميثانولية، لاستخدام في تحضير حبيبات الفضة النانوية الاستخلاص ب الساكسوليت هو الأسلوب المستخدم في تحضير خلاصة حشيشة الملاك الميثانولية، لاستخدام في تحضير حبيبات الفضة النانوية الملك والتي تم التحري عنها بالمجهر الالكتروني، تم الكشف عن ٥ مجاميع كيميائية نباتية،تقبيم النشاط المضاد الملاك، بالإضافة إلى أن اختبار الكشف المجاميع الكيميائية النباتية كان موجبًا للقلويدات والفلافونويد والعفس والستيرويدات والكومارين، حجم الملاك، بالإضافة إلى أن اختبار الكشف المجاميع الكيميائية النباتية كان موجبًا للقلويدات والفلافونويد والمناد المضاد البكتريا لما مالمالين المال الحبيبات التي اكتشافه المجهر الالكتروني كان ٩٠ ١٣. الميثانولي تعززت بو اسطة حبيبات الفضة النانوية. عرص ٢٠ ١٣. ١٣. متوسط حجم ± انحراف معياري. ان النشاط المضاد الميناط الميثانولي تعززت بو اسطة حبيبات الفضة النانوية. عرصت تنائج هذه الدر اسة ان لحبيبات الفضة النانوية المضاء منبعة لسيدم كمي كي ميوير وا

الكلمات المفتادية: نشاط مضاد للميكروبات، حشيشة الملاك، خلاصة ميثانولية، حبيبات الفضة النانوية.

## Introduction

The rise in the multi-drug resistant bacteria numbers due to wrong diagnosis and antibiotics' abuse is a global concern, and the pharmaceutical platform is always in need of new antibiotics to overcome the resistant species, in this manner, the plants contribute to this field with several entities and formulation such as in forms of extracts or nanoparticles<sup>(1,2)</sup>.

A traditional herb native to Indian subcontinent and applied widely in traditional medicine is the Himalayan*Angelica* family (Apiaceae) with more than 60 discovered species; the three most common species of *Angelica*, namely *Angelica glauca*, *Angelica cyclocarpa*, and *Angelica archangelica*<sup>(3)</sup>.

\*Corresponding author E-mail: pharm.zahraa@uomosul.edu.iq Received: 12/ 8/ 2022 Accepted: /15/ 1/2023

Iraqi Journal of Pharmaceutical Sciences

For centuries *Angelica glauca*, called Chora, has been used as an over-the-counter herbal preparation owing to its anti-inflammatory, diuretic, antispasmodic, laxative, and expectorant properties, and in the treating with respiratory and urinary tract infections<sup>(4)</sup>.

The nanotechnology has many procedures in nanoparticles synthesis; however the most popular one is the use of eco-friendly renewable medicinal plant extracts in green nanoparticle synthesis owing to its content of phytochemicals that act as reducing agents in capping and stabilizing nanoparticles. *Angelica* species contain several active metabolites like coumarins, flavonoids, tannins, alkaloids, and carbohydrates which possess antimicrobial and antioxidant properties<sup>(3)</sup>. Silver nanoparticles are the most common nanoparticles type in use due to the antiseptic activity owing to their antimicrobial and antifungal effects, adds an additive action to nanoparticles<sup>(5)</sup>.

The present work aims to synthesize Angelica glauca silver nanoparticles from its methanolic extract, detect phytochemicals, investigate the morphology in Angelica glauca AgNPs by using SEM, and finally in vitro antibacterial activity assessment of Angelica glauca of crude root extract and compare it with the silver nanoparticles forms.

## **Materials and Methods**

#### Extraction of the crude plant material.

The roots of *Angelica glauca* was purchased from India and manually pulverized to obtain 250 gram of dried fine powder. The dried root powder was subjected to soxhelation with one liter of 70% methanol for 24 h, sieved through folded gauze, and filtered using Whatman filter paper No.1. The crude extract was recovered from the solvent by a rotary evaporator under vacuum and reduced temperature at 45<sup>o</sup>C and yielded yellow residue of 13.3g and kept at refrigeration. The preparing *Angelica glauca* methanolic extract was used as a reducing agent in preparation *Angelica glauca* AgNPs, in the upcoming phytochemical screening and antibacterial activity assessments<sup>(6)</sup>.

# *Angelica glauca* mediated green silver nanoparticles synthesis

To prepare *Angelica glauca* silver nanoparticles, 2 ml of silver nitrate solution (10 mmol/L) was added to an equal quantity of the crude extract in a drop-by-drop manner, and the mixture was heated to 55 °C for 60 min and vigorous stirring, the mixture color change was objectively observed <sup>(1)</sup>. *Angelica glauca* extract (yellow solution) changed to a brown colloidal solution after adding the AgNO<sub>3</sub> solution indicating *Angelica glauca* silver nanoparticles formation. Then, the formed solution was centrifuged at 8000 rpm for 10 min then, sediment was preserved in refrigeration<sup>(7,8)</sup>.

#### Screening the phytochemical components

Phytochemical screening tests were carried out to confirm the presence of alkaloids, tannins, flavonoids, saponin, steroids, coumarins, terpenoids, quinine, proteins, and carbohydrates in *Angelica glauca* methanolic crude extract <sup>(9,10,11)</sup>.

Alkaloids detected by the addition of one ml of Mayer's reagent mixed with 1ml of the extract. A yellow color formation confirms the presence of alkaloids. Terpenoids detected by the addition of two drops of chloroform was mixed with 1ml of 0.1N hydrochloric acid then 1ml of the extract was added to the mixture and heated for 2 minutes. A red-brown color formation confirms the presence of terpenoids. Tannins detected by the addition of three drops of 10% lead a acetate were added to 1ml of the extract. Yellow precipitate formation is a positive reaction of tannins. Carbohydrate detected by the addition of one ml of sulfuric acid was added to 1ml of the extract. A red ring formation is an indication of carbohydrate. Saponin detected by the addition of three drops of distilled water was added to 1ml of the extract with shaking. A foam formation indicates the presence of saponin. Coumarins detected by the addition of one ml of diluted 0.1N nitric acid was added to 1ml of the extract. A yellow precipitate confirms the formation of coumarins. Flavonoid detected by the addition of one ml of ferric chloride was added to 1ml of extract, brown color confirmed the presence of flavonoids. Ouinine detected by the addition of one ml of 10% sodium hydroxide, was added to 1ml of the extract. A red color formation confirms the presence of quinine. Protein detected by the addition of one drop of 0.1N nitric acid was added to 1ml of the extract. A red color confirms the presence of proteins. Steroids detected by the addition of one ml of chloroform mixed with one ml 0.1N sulfuric acid were added to 1ml of the extract with shaking. The red color confirms the presence of steroids.

# The Angelica glauca AgNPs morphology detection

#### Scan electron microscope (SEM)

One drop of *Angelica glauca* AgNPs was placed on an SEM sample holder and air-dried. A SEM was performed by (TESCAN-MIRA 3, Czech Republic) to observe the morphology of *Angelica glauca* AgNPs <sup>(12)</sup>.

#### Antibacterial activity

The antibacterial assay for both the crude methanolic extract and the previously synthesized AgNPs derivative were carried out via disc diffusion method against the selected bacterial isolates of Staphylococcus aureus ATcc 3100, Bacillus subtilis ATcc 1100, Escherichia coli ATcc 7312, Klebsiella pneumonia ATcc 7012 obtained from microbiology laboratory, department of Clinical Laboratory Science. Subcultures were obtained after inoculating the isolated bacteria on a selective agar medium for each bacterial strain, the streaked fresh cultures were maintained in incubated for 24 h. at 37°C; on the other hand, activity of 1% dimethyl sulphoxide DMSO had been excluded by testing the absence of antibacterial activity against all isolated bacteria used in this study (negative control via disc diffusion method)<sup>(13)</sup>.

#### Disc diffusion method

Disc diffusion protocol was carried to determine the antibacterial activity of Angelica glauca crude methanolic fraction and the AgNPs derivative against S. aureus, B. subtilis, E. coli, and K. pneumonia. The susceptibility test was carried out by taking standardized inoculums of the bacterial cultures after adjusting its turbidity to achieve 0.5 McFarland then a swab from the bacterial culture was streaked on a Muller-Hinton agar medium plates. Six millimeters discs of Whatman filter paper No.1 were placed on inoculated agar medium after sterilization and soaking in Angelica glauca methanolic extract and Angelica glauca AgNPs separately in the following concentrations of 0.5, 1, 2, 4, 8, and 16 mg/ml after dissolving with negative control of 1% DMSO<sup>(14)</sup>.

Chloramphenicol 10 mg/disc was used as a positive control against all bacterial isolates; then, the inoculated plates were incubated at 37°C overnight. The inhibition zones were measured in millimeters against all the tested bacteria, susceptibility test carried out in triplicate, then mean and stander deviation were calculated<sup>(1)</sup>.

## **Results and Discussion**

The yield of the total crude extract after drying was 13.3g (5.3%) yielded from 250 g. However *Angelica* glauca AgNPs yield from methanolic extract was 1.5 g(0.6%).

Throughout history medicine utilized herbs which were considered safe, less toxic, and economic. The pharmacological activities displayed by medicinal plants are attributed to their phytochemicals used to cure several health conditions, and were the source of many medications in clinical practice <sup>(13)</sup>.

The plant extract yield in this work is dependent on several factors such as time of harvesting, geographical location, and extraction methods <sup>(15)</sup>.

On the other hand, the color change of the methanolic crude extract from yellow to dark brown color with colloidal dispersion after adding AgNO<sub>3</sub> which indicates the formation of *Angelica glauca* AgNPs. Nevertheless, the successful procedure in formations of AgNPs colloidal particles and color change to brown due to the reduction of Ag<sup>+</sup> to Ag<sup>o</sup> as a result of surface plasmon trembling in the methanolic extract<sup>(16)</sup>.

#### Phytochemical tests

The phytochemical qualitative test results based on color change and attributed to the interaction between the reagent and the various types of the phytochemicals. However phytochemical reactions of *Angelica glauca* roots were positive for alkaloids, flavonoids, tannins, steroids and coumarins. Furthermore, detection of terpenoids, carbohydrate, saponin, quinine, and proteins showed negative result.

Previously, phytochemicals obtained from *Angelica* glauca methanolic root which showed positive results for alkaloids, flavonoids, glycosides, steroids, saponin, and tannins, additionally, variation in the phytochemical profile may occur depending on the *Angelica* species, and the type of the diluent used for the extraction. <sup>(17,18)</sup>.

TT C	D 1		
Type of	Result		
phytochemicals			
Alkaloids	+		
Terpenoids	-		
Flavonoids	+		
Tannins	+		
Carbohydrates	-		
Saponin	-		
Quinines	-		
Proteins	-		
Steroids	+		
Coumarins	+		

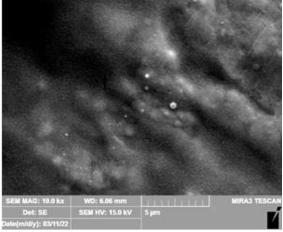
Table 1. The results of phytochemical screeningtests of Angelica glauca methanolic extract.

#### The detection test of Angelica glauca AgNPs Scan Electron Microscopy (SEM)

The size and morphology of derivatized *Angelica glauca* AgNPs were determined by SEM. Based on the SEM analysis, the *Angelica glauca* AgNPs size was  $70.9\pm13.60$  (mean  $\pm$ SD) in all measurements, Figure 1 (A, B).



A - Angelica glauca AgNPs size



B-Angelica glauca AgNPs shape

Figure 1. (A, B). Morphology (size and shape) of *Angelica glauca* AgNPs by SEM.

Nanoscience was globally used to produce a wide range of products. Although green synthesis of AgNPs using plant extracts to avoid toxicities, nowadays plant extracts are used widely in nanoparticles synthesis due to the presence of large quantities of phytochemicals which act as reducing agents to produce nanoparticles from metal salts without having any toxic by-product <sup>(19)</sup>.

The latter test was SEM which determined *Angelica* glauca AgNPs morphology in spherical particles, well dispersed, and their size was70.9 $\pm$ 13.60 (mean size in nm  $\pm$  SD), although previously, the AgNPs expressed spherical shape, with an average size of 21 nm in diameter. Data obtained from SEM with spherical particles and sizes less than 100 nm indicate a suitable nanoparticle formulation procedure <sup>(20)</sup>.

#### Antibacterial activity

In vitro antibacterial activity of Angelica glauca methanolic extract and AgNPs were assessed via the disc diffusion method, and their antibacterial activity was increased in a dose-dependent manner. Table 2 and Figure 2 demonstrate the antibacterial activity by measuring the inhibition zones expressed by Angelica glauca methanolic extract against G+ bacteria as S. aureus and B. subtilis were ranged from  $(0\pm0.00-0\pm0.00)$  and  $(0\pm0.00, 21\pm1.00)$ however, the inhibition zone against G- bacteria as E.coli, and K. pneumonia were ranged from (0.±0.0-10.6±1.15), and (0±0.00-0±0.00). The observed antibacterial activities of Angelica glauca AgNPs are illustrated in Table 3 and Figure 3; the inhibition zones against G+ such as S. aureus, and B. subtilis ranged  $(0.\pm 0.0-14.6\pm 0.57)$ and (14.6±0.57- $21.3\pm1.15$ ) respectively. Furthermore, the inhibition zones against G- bacteria as E. coli, and K. *pneumonia* ranged (10±1.00-16.3±1.15) and  $(0\pm 0.00-10.3\pm 0.57)$ , respectively.

**Table 2.** The inhibition zones' diameters in millimeters expressed by Angelica glauca crude methanolic extract against selected pathogens.

Conc.	S. aureus	B.subtilis	E .coli	K .pneumonia		
	Zone of inhibition (mean diameter in nm±SD)					
0.5	0±0.00	0±0.00	0±0.00	0±0.00		
1	0±0.00	0±0.00	10±0.00	0±0.00		
2	$0\pm0.00$	0±0.00	10±0.00	0±0.00		
4	$0\pm0.00$	13.3±1.52	10±1.00	0±0.00		
8	$0\pm0.00$	15.3±0.57	10±1.00	0±0.00		
16	$0\pm0.00$	21±1.00	10.6±1.15	0±0.00		

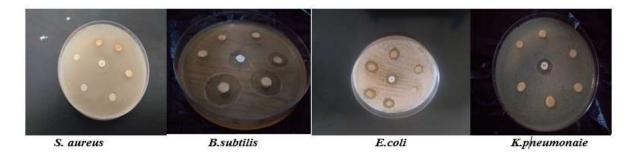


Figure 2. Inhibition zones displayed by testing the antibacterial activity of *Angelica glauca* crude methanolic extract against selected bacterial strains using disc diffusion assay.

Table 3. The inhibition zones' diameters in millimeters expressed by *Angelica glauca* methanolic extract derived-AgNPs against selected pathogens.

Conc.	S. aureus	B. subtilis	E. coli	K. pneumonia		
	Zone of inhibition (mean diameter in nm±SD)					
0.5	0±0.00	14.6±0.57	10±1.00	0±0.00		
1	0±0.00	14.6±1.52	11±1.0	0±0.00		
2	0±0.00	15±1.00	14.6±0.57	0±0.00		
4	13±1.00	15±0.00	15.3±0.57	0±0.00		
8	14±1.00	17±1.00	15.3±0.57	10.3±0.57		
16	14.6±0.57	21.3±1.15	16.3±1.15	10.3±0.57		

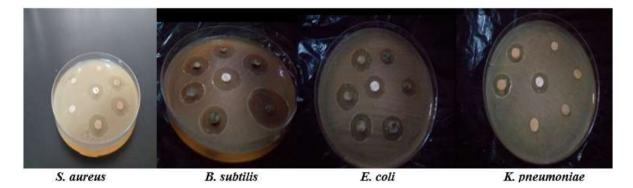


Figure 3. Inhibition zones displayed by testing the antibacterial activity of *Angelica glauca* methanolic extract derived-AgNPs against selected bacterial strains using disc diffusion assay.

Notably, the antibacterial activity of Angelica glauca in two formulations of methanolic and AgNPs showed that Angelica glauca AgNPs had higher antibacterial activity compared to the crude methanolic extract against all the tested bacteria. However several mechanisms of antibacterial action of phytochemicals have been reported, for instance they may act by inhibiting microbial growth, inducing cellular membrane perturbations, interference with certain microbial metabolic processes and modulation of signal transduction. Molecular basis for the modes of phytochemicals-based action in in vitro antimicrobial activity was the most accepted theory (21).

Angelica glauca crude methanolic extract inhibited B. subtilis > E. coli > S. aureus and K. pneumonia, while the last two bacteria showed resistance. Furthermore, AgNPs demonstrated more activity compared to its extract which were in the following manner *B. subtilis* > *E. coli* > *S. aureus*> *K. pneumonia.* The antibacterial effect of *Angelica glauca* AgNPs compared to its non-derivatized extract form is attributed to the small particle size and larger surface-to-volume ratios, in addition to coating phytochemicals with AgNO<sub>3</sub> will enhance the binding to bacterial cell membrane, which verifies bacterial cell death <sup>(1,22,23)</sup>.

## Conclusion

The two Angelica glauca formulas exhibited various degrees of antibacterial properties; Angelica glauca contain several healthrelated bioactive phytochemical compounds which can be used in curing health-related problems, however reducing phytochemicals into nanoparticles have several properties in pharmaceutical applications as well as our results.

### Acknowledgement

The authors acknowledge the College of Pharmacy–University of Mosul for providing the necessary facilities to carry out this study.

## **Conflict of Interest**

None

Funding

None

#### **Ethics Statements**

No animal or human include in the study.

## Author Contribution

The authors contribute in result analysis, writing and reviewing the manuscript.

#### References

- Riaz M, Altaf M, Ayaz M, Sherkheli MA, Islam A. Antibacterial and antioxidant potential of biosynthesized silver nanoparticles using aqueous root extract of *Angelica glauca*. Inorganic and Nano-Metal Chemistry. 2020 21:1-7.
- 2. China Medical Commission, Chinese Pharmacopeia Commission, Pharmacopoeia of the People's Republic of China (Part I) China Medical Science and Technology Press, Beijing, China, 2010.
- **3.** Sarker SD,Nahar L. Current Medicinal Chemistry. Bentham Science Publishers. 2004; 11(11): 1479-1500.
- **4.** Irshad M, Shahid M, Aziz S, Ghous T. Antioxidant, antimicrobial and phytotoxic activities of essential oil of *Angelica glauca*. Asian Journal of Chemistry. 2011;23(5):1947-1951.
- 5. Chernousova S, Epple M. Silver as antibacterial agent: Ion, nanoparticle, and metal. Angew. Chem. Int. Ed. 2013; 52: 1636–1653.
- 6. Handa SS, Khanuja SPS, Longo G, Rakesh DD. Extraction technologies for medicinal and aromatic Plants. Inter Cent Sci high Tech Trieste. 2008. PP. 24.
- Reda M, Ashames A, Edis Z, Bloukh S, Bhandare R, Abu Sara H. Green synthesis of potent antimicrobial silver nanoparticles using different plant extracts and their mixtures. Processes 2019; 7(510): 1-14.
- 8. Logeswari P, Silambarasan S, Abraham J, Synthesis of Silver Nanoparticles Using Plants Extract and Analysis of their Antimicrobial

Property. J. Saudi Chem. Soc. 2015; 19: 311-317.

- **9.** Shome U, Joshi P, Sharma HP. Pharmacognostic studies on *Artemisia scoparia* Waldst and Kit. Proc. Plant Sci., 1984; 93:151-164.
- Perveen AN, Ijaz S, Ghaffar NA. Comparitive phytochemical and physicochemical study of seeds of the genus Angelica L. From neelum valley azad Kashmir, Pakistan. Pak. J. Bot. 2020; 52(1):257-60.
- **11.** Zaman KU, Bakht J, Munir IQ, Shafi M. Phtyto-chemical composition, antimicrobial and phyto-toxic activity of *angelica glauca* (apiaceae). Pakistan Journal of Botany. 2018;50(5):1893-8.
- **12.** Csakvari AC, Moisa C, Radu DG, Olariu LM, Lupitu AI, Panda AO, *et al.* Green synthesis, characterization, and antibacterial properties of silver nanoparticles obtained by using diverse varieties of Cannabis sativa leaf extracts. Molecules. 2021;26(13):4041.
- **13.** Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by standardized single disk method. Am. J. Clin. Pathol. 1966;45: 493-496.
- KHALEEQ-UZ-ZAMAN JB, Shafi M, Munir I. Phtyto-Chemical Composition, Antimicrobial and Phyto-Toxic Activity of Angelica glauca (Apiaceae). Pak. J. Bot. 2018;50(5):1893-1898.
- **15.** Vashistha RK, Butola JS, Nautiyal BP, Nautiyal MC. Phenological attributes of *Angelica glauca* and A. *archangelica* expressed at two different climatic zones in Western Himalaya. Journal of Medicinal and Aromatic Plants. 2010; 1(1): 7-12.
- 16. Dubey SP, Lahtinen M, Sillanpää M. Tansy fruit mediated greener synthesis of silver and gold nanoparticles. Process Biochemistry. 2010;45(7):1065-71.
- **17.** Arya P, Mehta JP, Maurya VK. Methanolic Extract of *Angelica glauca* Edgew Root and Stem: A Possible Component of Herbal Medicines against Respiratory Infections. Indian Journal of Pharmaceutical Education and Research. 2021;55(2):S552-62.
- Premanath R, Sudisha J, Devi NL, Aradhya SM. Antibacterial and antioxidant activities of fenugreek (*Trigonella foenumgraecum* L.) leaves. Res. J. Med. Plant., 2011; 5: 695-705.
- Siddiqi KS, Husen A, Rao RAK. A review on biosynthesis of silver nanoparticles and their biocidal properties. Journal of Nanobiotechnology. 2018; 16(14):1-28.

- **20.** Liu H-X, Suna S-Q, Lvb G-H, Chanb KKC. Study on *Angelica* and its different extracts by Fourier transform infrared spectroscopy and two-dimensional correlation IR spectroscopy. Spectrochimica Acta Part A, 2006; 64:321–326.
- **21.** Omojate Godstime C, Enwa Felix O, Jewo Augustina O, Eze Christopher O. Mechanisms of antimicrobial actions of phytochemicals against enteric pathogens–a review. J Pharm Chem Biol Sci. 2014; 2(2):77-85.
- **22.** Dhand V, Soumya L, Bharadwaj S, Chakra S, Bhatt D, Sreedhar B. Green synthesis of silver nanoparticles using *Coffea arabica* seed extract and its antibacterial activity. Materials Science and Engineering: C. 2016;58:36-43.
- **23.** Ahmed S, Ahmad M, Swami BL, Ikram S. A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise. J Adv Resear. 2016;7(1):17-28.



This work is licensed under a Creative Commons Attribution 4.0 International License.