DOI: https://doi.org/10.31351/vol33iss1pp94-103

Ultrasound-Assisted Extraction and Cytotoxic Activity of Fennel Leaves: Process Optimization and Thin Layer Chromatography

Thukaa Z. Abdul-Jalil *,1

¹Department of pharmacognosy and Medicinal plants, collage of pharmacy, University of Baghdad, Baghdad, Iraq.

Abstract

Foeniculum vulgare commonly known as fennel belonging to the family Umbelliferae, is cultivated throughout the world as a spices as well as medicinal plant possessing antimicrobial, antidiabetic, antioxidant, anti-inflammatory and others. A Single Factor Experiment analysis has been employed in this study to optimize caffeic acid extraction from fennel leaves by ultrasound assisted extraction method in which three variables single factor examined which were ethanol concentration, extraction time and ultrasound frequency. The study's objective was to assess the effects of variables chosen on caffeic acid's extraction performance, preliminary evaluated by thin layer chromatography (TLC) and evaluate the cytotoxic effect of optimized ethanolic extract of this plant on human breast cancer (MDA-MB-468) cell line. The experimental results revealed that the most influential extraction variables were: 80% ethanol as extracting solvent, 15 min was the time to achieve the good extraction efficacy and 40 KHz ultrasound frequency and The IC50 value of the cytotoxic effect (P < 0.05) of optimized ethanolic extract of Iraqi Foeniculum vulgare leaves on MDA-MB-468 cell line after 72 h of incubation were significantly high (46.89 µg/ml). The results of the study will be used to create a natural medicine for the symptoms of various diseases and to standardize the processes for extracting caffeic acid from natural items, particularly fennel.

Keywords: Caffeic acid, Cytotoxic activity, Fennel, Single factor experiment, Ultrasound assisted extraction.

استخلاص أوراق حبة الحلوة بمساعدة الموجات فوق الصوتية والنشاط السام للخلايا: العملية الأمثل و الفصل اللوني بالطبقة الرقيقة

ذكاء زهير عبد الجليل *١٠

اقسم العقاقير والنباتات الطبية, كلية الصيدلة, جامعة بغداد, بغداد, العراق.

الخلاصة

يُزرع Foeniculum vulgare المعروف باسم حبة الحلوة الذي ينتمي إلى عائلة الخيمية، في جميع أنحاء العالم كتوابل بالإضافة إلى نبات طبي يحتوي على مضادات الميكروبات ومضادات السكر ومضادات الأكسدة ومضادات الالتهابات، ... إلخ. تم استخدام تحليل تجربة العامل الواحد في هذه الدراسة لتحسين استخلاص حامض الكافيك من أوراق حبة الحلوة عن طريق وسيلة الاستخلاص بمساعدة الموجات فوق الصوتية حيث تم فحص ثلاثة متغيرات عامل واحد وهي تركيز الإيثانول ووقت الاستخلاص وتردد الموجات فوق الصوتية. كان الغرض من هذه الدراسة هو تقييم تأثيرات المتغيرات المختارة على أداء استخلاص حامض الكافيك، وتقييم أولي بواسطة الفوني بالطبقة الرقيقة (TLC)وتقييم التأثير السام للخلايا للمستخلص الإيثانولي الأمثل لهذا النبات على سرطان الثدي البشري (MDA-MB-468)خط الخلية. أظهرت النتائج التجريبية أن أكثر متغيرات الاستخلاص تأثيرًا كانت: ٨٠٠٪ إيثانول كمذيب استخلاص، و١٥ دقيقة هي الوقت المناسب لتحقيق كفاءة استخلاص جيدة وتردد الموجات فوق الصوتية ٥٠ كيلو هر تز وكانت قيمة 100 للتأثير السام للخلايا (٢٥٠ ٥٠) للمستخلص الإيثانولي الأمثل لأوراق حبة الحلوة العراقية على خط خليا هما الموجات الطبيعية وخاصة حبة الحلوة وعمل علاج طبيعي ضد أعراض الأمراض المختلفة. المتناط السام للخلايا، حبة الحلوة وعمل علاج طبيعي ضد أعراض الأمراض المختلفة. المسوتية. الكلمات الرئيسية: حامض الكافيك، النشاط السام للخلايا، حبة الحلوة، تجربة العامل الواحد الاستخلاص بمساعدة الموجات فوق الصوتية.

Introduction

Plants have been shown to have significant concentrations of a number of secondary metabolites, such as phenolic compounds, flavonoids, carotenoids, terpenoids and alkaloids which serve as nutrients, antimicrobials and antioxidants that can be utilized to cure and prevent a wide range of human ailments ⁽¹⁾. They are also crucial to the defensive mechanism of the plant; Particularly, polyphenolic compounds which are the most prevalent bioactive components found in edible plants. These compounds are important

sources of metabolites that promote human health because they act as non-enzymatic antioxidants in cells to stop damage from oxidative stress and reduce the risk of developing various chronic diseases including cancer ⁽²⁾. For these naturally occurring bioactive compounds to be successfully obtained, extraction methods must be carefully examined, there have been several suggested techniques for isolating natural bioactive substances ⁽³⁾

*Corresponding author E-mail: aljas_dadid@yahoo.com

Received: 18/12 / 2022 Accepted: 5/4/2023

Iraqi Journal of Pharmaceutical Sciences

Traditional techniques including maceration, percolation, Soxhlet extraction, reflux extraction, and infusion are frequently employed, but they take a long time and require a lot of efforts, furthermore, extracting solvents are not ecofriendly, so in order to address these issues, ecofriendly alternatives have recently been developed. (4) Examples of these eco-friendly procedures include supercritical fluid extraction, microwaveassisted extraction. and ultrasonic-assisted extraction (4,5)

Due to its usefulness and affordability, ultrasonic-assisted extraction (UAE), among other things, is very promising for the food and herbal industries, also enables for low-temperature extraction, uses less time and energy and preserves the extract's quality. (6) UAE extracts bioactive compounds from its natural sources using high-intensity sound waves in which plant interstitial tissues are damaged by the sonic cavitation produced by UAE, increasing the solvent-plant material contact area leading to a higher extraction yield of bioactive compounds compared with traditional extraction methods, saving time, cost and labor (7).

This research focuses on evaluating fennel's anticancer properties and using the UAE approach to extract phenolic components, particularly caffeic acid; due to its flavor, *Foeniculum vulgare*, a medicinal plant in the Umbelliferae (Apiaceae) family, has been utilized and known to humans since ancient times and was grown in practically every nation ⁽⁸⁾. It has more than 100 names in addition to the common name "fennel" with a lengthy history of usage as medicine and is a well-known and traditional herb. ⁽⁹⁾ Studies have shown that *F. vulgare* is capable of efficiently treating a wide range of infectious diseases with bacterial, fungal, viral, mycobacterial and protozoal origins ⁽⁹⁻¹¹⁾.

It has hepatoprotective (12), cytoprotective (13), antitumor (14), anticancer (14) and hypoglycemic effects (15). According to Koppula S. and Kumar H., *F. vulgare* can lower stress and has a unique memory-enhancing function (16). Limited clinical trials and animal studies indicate that long-term usage of *F. vulgare* is not hazardous. (13) Fennel can be eaten on a regular basis in a variety of ways, including fresh in salads and snacks, cooked in stews, boiled, grilled or baked foods or even used to make herbal teas or alcoholic beverages. Due to its

valuable nutritional constituents rich in polyphenolic chemicals, a diet containing the reasonable amounts of fennel may have positive effects on health (17).

Through the use of single-factor experiment analysis, preliminary chemical tests, thin layer chromatography (TLC), and cytotoxic activities that were specifically examined to validate the ideal UAE conditions for fennel leaves, this research makes a significant contribution by being the first to evaluate the application of the UAE technique to extract biologically active compounds like caffeic acid from fennel leaves crude extract, one of the plants that are most frequently used in traditional medicine and foods.

Materials and Methods

Plant Collections

Foeniculum vulgare leaves were procured in October 2021 from local markets in Baghdad/Iraq and then verified in the Department of Pharmacognosy and Medicinal Plants /College of Pharmacy/ University of Baghdad. The leaves were then subjected for series of workup, water-washed, shade-dried for two weeks at room temperature then grounded with electric blender, weighted and processed to the extraction techniques.

Experimental work

The experimental work was divided into sequential steps:

Extraction of plant material

Sixty grams of the powdered plant material was extracted by maceration in n-hexane for 2 days with frequent shaking, at room temperature, the extract was filtered off, this procedure was repeated three times for defatting purpose (18), then the plant material was left to dry and subjected to extraction by Ultrasound assisted extraction (UAE) method that was done by using probe ultrasonicator with ethanol as a solvent. The UAE was carried out under the following experimental conditions:

Plant material is directly in touch with ultrasonic waves in a probe ultrasonicator at a temperature of 25 °C, for 10 minutes, with a solvent to solids ratio of 10:1 ml/gm, 60% ethanol as extracting solvent and a sonication frequency of 40 Hz. ⁽¹⁹⁾. Later on, the crude ethanol extract was filtered and evaporated to dryness using a rotary evaporator, as shown in Figure 1.

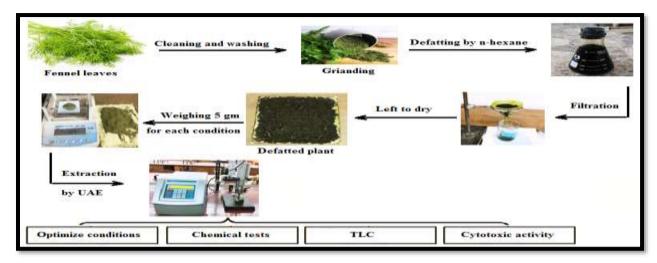


Figure 1. Illustration graphic for extraction procedure of Iraqi Foeniculum vulgare leaves.

Experimental Design using Single Factor Experiment

The impact of each factor on fennel extraction was examined using a single-factor experiment. The other special factors remained the same, but one of them altered. Four factors (solvent/solid ratio, ethanol concentration, time of extracting, and ultrasound frequency) were reported to be different for similar phytoconstituents from different plant varieties; for all of the above a single factor experiment was chosen each time; the level gradients of each factor were: solvent/solid ratio was preset as 10:1 ratio since the study was done in small scale; ethanol consternations were 60%,70%, and 80%; extracting times were 5,10, and 15min; ultrasound frequencies were 20,40, and 60KHz (2,19).

Preliminary phytochemical examination of crude extract

The dried extract from crude ethanolic extract had been subjected to preliminary phytochemical assay for initial screening of possible phytochemicals applying chemical tests for detecting polyphenols and flavonoids in the extracts (20,21).

Test for phenolic

One milliliter of crud ethanolic extract was dissolved in 1 ml of 5% ferric chloride, the color was observed, a deep green to black coloration indicates the presence of phenolic acids. (20)

Test for flavonoids

One milliliter of crud ethanolic extract was dissolved in 2 ml 1% potassium hydroxide in a test tube, and the color was observed. A yellow color indicates the presence of flavonoids. (21)

Qualitative identification by thin layer chromatography

The optimized ethanolic extract of Iraqi Foeniculum vulgare leaves were subjected to thin layer chromatography for the qualitative estimation of phenolic acid (caffeic acid). This was done using ready-made aluminum silica gel plates GF 254, detection by UV light detector at 254nm and 366nm

wave lengths, standard caffeic acid (sigma-aldrich, USA), and five different solvent systems for identification. (22,23):

- S₁: Toluene: ethyl acetate: formic acid (9: 3: 1.25)
- S₂: Ethyl acetate: toluene: formic acid: methanol (3: 3: 0.8: 0.2)
- S₃: Toluene: dioxan: acetic acid (9.2: 4:0.6)
- S4: Toluene: chloroform: acetone: formic acid (8: 4: 3: 3)
- S5: Chloroform: acetone: formic acid (75:16.5:8.5)

Cytotoxic assay method

On the viability of the human MDA-MB-468 cancer cell line, an optimal ethanolic extract of Iraqi Foeniculum vulgare leaves was tested using the MTT colorimetric assay. (24). The human MDA-MB-468 breast cancer cell lines were donated by ATCC. They were kept in the cell bank of the tissue culture research center at College of Pharmacy/ University of Al-Mustansiriyah. The cells were placed in liquid nitrogen storage, frozen at (80 C°, 24 h), thawed at 37 Co, and then given 10 ml of fresh media. The cells were obtained using centrifugation. After that, the cells were moved into a flask of 75 cm² and cultivated in 25 ml of fresh medium. A549 cells were maintained in 500 cc of RPMI-1640 Medium. Penicillinstreptomycin-amphotericin B100X as an antiseptic and 50 ml of 10% FBS from complete medium were administered as supplements. Cells were cultivated in flasks with a 75 cm² surface area. Incubated at 5% CO₂ and 95% humidity was (37 C°).

Flasks containing A549 cells were passaged under sterile conditions as soon as the cells reached 90% confluence. PBS 5ml solution was used to wash the cells. After that, cells were incubated in a trypsin solution at 37 C° for two minutes to allow them to separate from the flask's bottom. A conical tube was then filled with the cell suspension and an equivalent volume of complete growth medium (50ml). The cells were also centrifuged (3 min, 1200 rpm). The cell pellet was then resuspended in enriched growth media

after the supernatant was discarded (fresh). In order to calculate the number of cells needed, a haemocytometer was employed under a microscope (25)

Cell Viability by MTT Colorimetric Assay

The vitality of MDA-MB-468 cancer cells was examined using the MTT test in order to determine the effects of the optimized ethanolic extract of Iraqi Foeniculum vulgare leaves. In 96-well flat-bottom tissue culture plates, cell suspensions (100 ul) were added at concentrations of 5 x 10³ cells per well and incubated for 24 hours under standard conditions, 4 x 10³ cells for 48 hours, and 3 x 10³ cells for 72 hours. After 24 hours, the cells were treated with various concentrations of the optimal (80% ethanol as extracting solvent, solvent to solids ratio of 10:1 ml/gm, temperature of 25 °C with 15 min as the time to achieve the good extraction efficacy and 40 KHz ultrasound frequency) ethanolic extract of Iraqi Foeniculum vulgare leaves (0.15, 0.35, 0.75, 1.52, 3.125, 6.25, 12.5, 25, 50 and 100 µM).

The cell culture medium was removed after a recovery period of 24 hours, 48 hours, and 72 hours, and cultures were then incubated for 4 hours at 37 C° with a medium containing 30 μ L of MTT solution (3 mg/ml of MTT powder in PBS). This medium was removed after 4 hours by gently inverting it and tapping on paper. To the control wells, just 100 μ L of growth medium were introduced. Then, 100 μ L of DMSO were added to each well, which were then left in the dark at room temperature for about (15-20 min) By utilizing a multi-scan reader with a wavelength of 540 nm to detect each well's absorbance and 650 nm to account for background absorption, the absorbance of each well was calculated $^{(24,26)}$.

The percentage of cytotoxicity, or the rate at which cell growth is inhibited, was estimated as follows:

[Percentage Inhibition Rate= (A-B/A) *100] Where [A, and B] denoted, respectively, the optical densities of the control and tested substances.

Determination of the Half-Maximal Inhibitory Concentration (IC_{50})

By creating a dose-response curve and analyzing the impact of various antagonist concentrations on reversing agonist activity, it is possible to establish the IC₅₀ of the optimized ethanolic extract of Iraqi Foeniculum vulgare leaves. By figuring out the concentration required to inhibit 50% of the maximum biological response of the agonist, the IC₅₀ values for a particular antagonist can be derived. The environment in which IC₅₀ values are measured has a significant impact on those results. In general, agonist activity will be decreased more as inhibitor concentration increases. As agonist concentration rises, IC₅₀ value rises. Furthermore, several factors may affect the IC₅₀ value depending on the kind of inhibition. The IC₅₀ shows the concentration of the optimum ethanolic necessary for a 50% suppression of cell viability, according to the in vitro MTT test.

Based on the results of the in vitro MTT experiment, the optimum ethanolic IC_{50} values were determined 72 hours after the cells were exposed to the compounds. 1.52–100 μM concentrations were used $^{(27)}$.

Data Analysis

The statistical analysis of the optimized ethanolic extract of Iraqi *Foeniculum vulgare* leaves on MDA-MB-468 cancer cell was performed using the nonlinear regression curve fitting software Graph Pad Prism 8.1. One-way ANOVA with Post hoc Tukey was used to compare all groups within a single MTT plate Statistics were judged significant at p < 0.05

Results and Discussions

Extraction outcome

During last decades, aromatic plants widely used in food as herbs and spices, are extensively studied for their phenolics and volatile compounds. aromatic herbs and spices, Among (Foeniculum vulgare) has a long history of use, both for its sensory and its functional (carminative, stomachic, antispasmodic and antimicrobial) properties (28). In spite of the importance of Foeniculum vulgare as an aromatic herbs and spices, the phytochemical studies concentrated on the volatile oils secondary metabolites especially fennel transanethole, estragole, fenchone, and limonene (29). Therefore, the chosen of phenolic compounds especially caffeic acid in this study is the main target have different pharmacological because they activities.

In this study, the first step involved as much fat removal as possible by maceration with n-hexane then the extraction was done by Ultrasound assisted extraction (UAE) probe type which is a quick, safe, and environmentally responsible method of extracting natural compounds that yields high-quality extracts (2,5). The findings revealed that probe UAE produced the higher percentage yield in the Iraqi Foeniculum vulgare leaves after each single optimizing factor tested with different variables so as to get to the best yielding condition for it, then they were summited with each other to verify the optimized condition (80% ethanol as extracting solvent, solvent to solids ratio of 10:1 ml/gm, temperature of 25 °C with 15 min as the time to achieve the good extraction efficacy and 40 KHz ultrasound frequency) to get the higher extraction yield which equals to 2.78 gm.

Single factor experiment analysis

Due to the complexity of the herbals' chemical composition and the components' affinities towards various changeable parameters, (30) numerous experiments had been done to obtain an appropriate optimal condition. The UAE parameters were optimized using single factor experimental analysis.

Effects of ethanol concentration

superiority The hydro-alcoholic combinations over other common solvents can be explained by the fact that they contain water, which may expand the plant material and make it easier for the solvent to penetrate the solid matrix and improve extractability (31). Ethanol is the most efficient solvent for the recovery of phenolic acid (32) especially caffeic acid; target of this study, therefore ethanol/water was chosen as the extracting solvent. concentrations of ethanolic solutions (60% to 80%) were examined using a 10:1 solvent/solid ratio, 10 min of extraction time, and 40 KHz ultrasonic frequency in order to examine the influence of ethanol/water concentrations on extraction yield. Figure 2 shows the effect of the solvent on the fennel extraction, when the ethanol concentration increased, the extraction efficiency (percentage yield) gradually increased. Thus, 80% ethanolic solvent was applied in the later experiments.

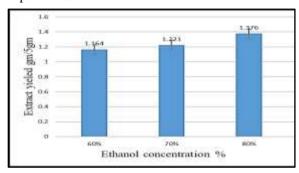


Figure 2. The effect of ethanol concentration on the extraction efficiency (percentage yield) of the fennel leaves extracts.

Effect of extracting time:

Time of extraction is an important parameter effecting on the extraction efficiency, in order to evaluate the influence of extraction time on Iraqi *Foeniculum vulgare* leaves, extractions were performed in the range of 5-15 min with 80% ethanol, 10:1 solvent/solid ratio and 40 KHz ultrasound frequency as shown in Figure 3, when the extracting time increased from 5 min to 15 min, the extraction efficiency (percentage yield) increased. The results showed that ultrasound could accelerate the release of bioactive constituents from the material when the ultrasound time is prolonged (33) Therefore, 15 min was chosen in the later experiments.

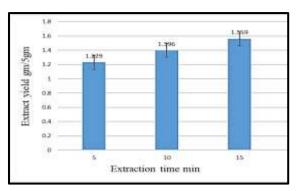


Figure 3. The effect of extraction time on the extraction efficiency (percentage yield) of the fennel leaves extracts.

Effects of Ultrasound frequency

Another crucial factor that affects extraction effectiveness is ultrasound frequency, the higher ultrasonic frequency applied, the more bubbles formed. (32) Every point of the extraction solution can be reached by ultrasonic waves of greater amplitude (ultrasonic frequency) which can improve the effectiveness of the extraction, while a very high frequency can disturb the plant in the extracts. (34) The effects of ultrasound frequency on fennel extraction in the range of 20 KHz to 60 KHz were examined with 80% ethanol, 10:1 solvent/solid ratio and 10 min extraction time, the results are presented in Figure 4. when the ultrasound frequency rose from 20 to 40 KHz the extraction efficiency increased. As the ultrasound frequency rose from 40 to 60 KHz, the extraction efficiency slightly decreased, as high frequency low power density generates cavitations with producing large number of reactive radicals like hydroxyl (OH) and hydrogen peroxide (H₂O₂) radicals causing the degradation of compounds. (35) Thus, 40 KHz was chosen as the preferred ultrasound frequency for the UAE process.

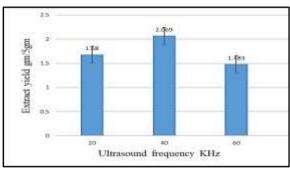


Figure 4. The effect of ultrasound frequency on the extraction efficiency (percentage yield) of the fennel leaves extracts.

Preliminary phytochemical examination of fennel extract:

After obtaining the aqueous ethanolic extract under optimized conditions, phytochemical screening assays were performed with the appropriate tests as they are simple, quick and inexpensive procedures. -Test for phenolic: a positive result was found as deep green to black coloration indicate the presence of phenolic acids.

-Test for flavonoids: a positive result was present a yellow color indicates the presence of flavonoids.

Qualitative evaluation by thin layer chromatography (TLC)

Thin layer chromatography (TLC) can be used on an analytical scale to track the course of a reaction and quickly determine how many components are in a combination, or on a preparative scale to purify small amounts of a product. The objective of TLC is to produce well-defined and well-separated spots that will support the identity of the chemical in the mixture when a compound's $R_{\rm f}$ is compared to the $R_{\rm f}$ of reference compound that is known to exist.

Thin layer chromatography of different aqueous ethanolic extracts obtained from *Foeniculum vulgare* leaves several trials conditions confirm the following:

A.The presence of numbers of phytoconstituents in different aqueous ethanolic concentrations (60%-80%) of *Foeniculum vulgare* leaves. One of these phytoconstituents, appeared as a single compact spot having the same color and R_f value as that caffeic acid standard on TLC plates which was identified by using three different developing solvent systems (S1, S2 and S3) and detected by using UV-light at 254nm and 366nm. as shown in Figure 7.

The $R_{\rm f}$ values of this spot and its corresponding standard (caffeic acid) in three developing solvent systems were calculated and listed in Table 1.

Table 1. The R_f values of the best three developing solvent systems.

Solvent systems	Caffiec acid standard	Ethanolic concentration 60%	Ethanolic concentration 70%	Ethanolic concentration 80%
S1	0.49	0.494	0.488	0.488
S2	0.827	0.827	0.814	0.82
S3	0.642	0.63	0.636	0.63

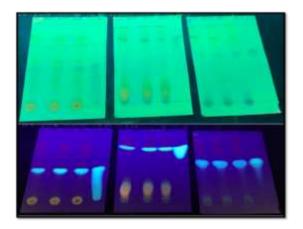


Figure 7: TLC chromatogram of aqueous ethanolic extract of fennel leaves at three different concentrations (60%,70%,80%) using silica gel GF_{254nm} as adsorbent and the three mobile phase S1, S2, S3 respectively, UV detection at 254 and 366 nanometers. S₁: toluene: ethyl acetate: formic acid (9: 3: 1.25), S₂: ethyl acetate: toluene: formic acid: methanol (3: 3: 0.8: 0.2), S₃: toluene: dioxan: acetic acid (9.2: 4:0.6)

B.The presence of numbers of phytoconstituents in aqueous ethanolic extracts of *Foeniculum vulgare* leaves at **different extraction times** (5,10,15 min). One of these phytoconstituents, appeared as a single compact spot having the same color and R_f value as that caffiec acid standard on TLC plates which was identified by using three different developing solvent systems (S1, S2 and S3) and detected by using UV-light at 254nm and 366nm. as shown in Figure 8.

The $R_{\rm f}$ values of this spot and its corresponding standard (caffiec acid) in three developing solvent systems were calculated and listed in Table 2.

Table 2. The R_f values of the best three developing solvent systems.

Solvent systems	Caffiec acid	Etraction time	Etraction time	Etraction time
	standard	5 min	10 min	15 min
S1	0.462	0.447	0.46	0.458
S2	0.731	0.725	0.725	0.737
S3	0.532	0.536	0.528	0.53

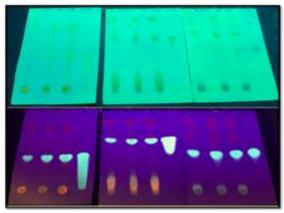


Figure 8: TLC chromatogram of aqueous ethanolic extract of fennel leaves at three different extraction times (5,10,15 min) using silica gel GF_{254nm} as adsorbent and the three mobile phase S1, S2, S3* respectively, UV detection at 254 and 366 nanometers. S₁: toluene: ethyl acetate: formic acid (9: 3: 1.25), S₂: ethyl acetate: toluene: formic acid: methanol (3: 3: 0.8: 0.2), S₃: toluene: dioxane: acetic acid (9.2: 4:0.6)

C. The presence of numbers of phytoconstituents in aqueous ethanolic extracts of *Foeniculum vulgare* leaves at different **ultrasound frequency** (20, 40, 60 KHz). One of these phytoconstituents, appeared as a single compact spot having the same color and $R_{\rm f}$ value as that caffeic acid standard on TLC plates which was identified by using three different developing solvent systems (S1, S2 and S3) and detected by using UV-light at 254nm and 366nm. as shown in Figure 9.

The $R_{\rm f}$ values of this spot and its corresponding standard (caffeic acid) in three developing solvent systems were calculated and listed in Table 3.

Table 3. The R_f values of the best three developing solvent systems.

Solvent systems	Caffiec acid	Ultrasound	Ultrasound	Ultrasound
	standard	frequency 20 KHz	frequency 40 KHz	frequency 60 KHz
S1	0.468	0.456	0.46	0.46
S2	0.667	0.66	0.654	0.655
S3	0.735	0.736	0.736	0.729

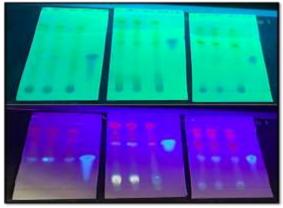


Figure 9.TLC chromatogram of aqueous ethanolic extract of fennel leaves at three different ultrasound frequency (20, 40, 60 KHz) using silica gel GF_{254nm} as adsorbent and the three mobile phase S1, S2, S3 respectively, UV detection at 254 and 366 nanometers. S₁: toluene: ethyl acetate: formic acid (9: 3: 1.25), S₂: ethyl acetate: toluene: formic acid: methanol (3: 3: 0.8: 0.2), S₃: toluene: dioxane: acetic acid (9.2: 4:0.6)

From the above, caffeic acids TLC qualitative evaluation results can run in the same path with the Single factor experiment analysis of UAE results proposing that the optimized UAE is a simple, trouble-free and effective option for recovering and assessing the quality of phenolic

acids, particularly caffeic acid recovered from fennel species leaves.

Cytotoxic assay

The optimized ethanolic extract of Iraqi Foeniculum vulgare leaves, obtained by probe UAE method as it gave the highest concentration of the bioactive constituents (caffeic acid) were monitored by TLC, was subjected to the cytotoxic assay for the first time on human tumor cell line; pleural effusion breast metastatic adenocarcinoma MDA-MB-468 cells by the MTT test.

With a maximum concentration cytotoxic activity at 100 µg/ml and a minimum concentration cytotoxic activity at 0.15 µg/ml, Figure 9 illustrated the cytotoxic effect of the optimized ethanolic extract on the MDA-MB-468 cell line and demonstrated that the cytotoxic effects of the optimized ethanolic extract had a concentration and time-dependent pattern. Its cytotoxic effect on MDA-MB-468 breast cancer cell lines was 28.37% at a dosage of 12.5 µg/ml after 48 hours of incubation. With a rise in the optimal ethanolic extract concentration and incubation period, the cytotoxic effects were more pronounced. As a result, the cytotoxic effect on the MDAMB- 468 breast cancer cell lines after 72 hours of incubation was 72.64% on a concentration of 100 μg/ml.

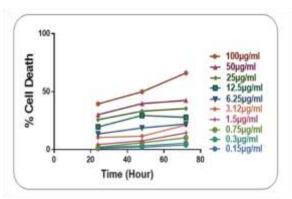


Figure 9. Response curve at different conc.(100-0.15µg/ml) of optimized ethanolic extract of Iraqi *Foeniculum vulgare* leaves, on MDA-MB-468 cell line after 24hr,48hr and 72 hr.

The cytotoxic effects of conventional medicinal compounds are categorized into four groups based on the IC₅₀ value. The classifications of very active, relatively active, weakly active, and inert cytotoxic chemicals are based on the IC50 values of 0-20, 20-100,100-1000, and >1000 μg/ml, respectively (24). On the MDA-MB-468 cell line, the optimized ethanolic extract of Iraqi Foeniculum vulgare leaves had a cytotoxic impact with an IC50 value of 46.89 g/ml. The optimized ethanolic extract of Iraqi Foeniculum vulgare leaves is categorized as a " relatively active" compound by the anticancer compound classification criteria. Plots of the concentrations of log-transformed Foeniculum vulgare extract were made to show the dosage response. Through the use of nonlinear regression analysis, IC50 values were calculated (Prism Pad 8.1). Results represent the standard error of the mean (SEM) for triplicate data p<0.05, as shown in Figure 10.

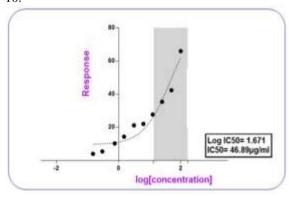


Figure 10. Dose-response curves of IC₅₀ for the optimized ethanolic extract of Iraqi *Foeniculum vulgare* leaves

According to the results of the phytochemical analysis, the optimized ethanolic *Foeniculum vulgare* leaves extract contained polyphenolic components with antioxidant and free radical scavenging activities. Polyphenolic components demonstrated an anticancer effect on a number of key factors, including cell proliferation,

metastasis and angiogenesis⁽³⁶⁾. Previous research showed that caffeic acid had an anti-proliferative and pro-apoptotic effect on the breast cancer MCF-7cell line ⁽³⁷⁾, also alters the expression of tumor cell cycle regulatory proteins in diverse ways to block the cell cycle at different stages and promote apoptosis in cancer cells ⁽³⁸⁾.

Conclusion and Perspectives

The findings demonstrate that the optimum UAE conditions for fennel extraction were 80% ethanol, 15 min. extraction time and 40 KHz ultrasound frequency; in addition to, the high cytotoxic potential of *Foeniculum vulgare* for *in vitro* manner, which is attributed mainly to the occurrence of caffeic acid and other related polyphenolic compounds that were detected using the MTT cytotoxic assay. Further investigations are needed to complete the phytochemical profile, pharmacology mechanisms and pharmacokinetics studies of the optimized ethanolic *Foeniculum vulgare* leaves extract.

Acknowledgment

Thanks for the College of Pharmacy/ University of Baghdad for their support to carry out this work and the authors are grateful to assist. prof. Dr. Salema Sultan Salman, College of Pharmacy/ University of Baghdad and to prof. Dr. Sukaena Abass Department of Biology / College of Science/ University of Baghdad for providing the scientific assistance for this work.

Conflicts of Interest

The authors declare no conflict of interest regarding the publication of the manuscript.

Funding

The research has not received financial support from any Institution.

Ethics Statements

The manuscript did not include human and/or animal studies, so ethical approval is not needed for this research.

Author Contribution

The author (Thukaa Zuhair Abdul-Jalil) planned the experiments, accomplished the planned procedures, including sample preparation, extraction process, phytochemical analysis by TLC, evaluation of cytotoxic activity, statistical analysis, and wrote the manuscript.

References

- 1. Mohammadinejad R., Shavandi A., Raie D. S. et al.: Plant molecular farming: production of metallic nanoparticles and therapeutic proteins using green factories. *Green Chemistry* 2019; 21(8): 1845–1865.
- 2. Ratananikom K and Premprayoon K: Ultrasonic-assisted extraction of phenolic compounds, flavonoids, and antioxidants from Dill (*Anethum graveolens* L.) *Scientifica* 2022; 2022(1):1-6

- **3.** Barba F. J., Zhu Z., Koubaa M., Sant'Ana A. S., and Orlien V.: Green alternative methods for the extraction of antioxidant bioactive compounds from winery wastes and byproducts: a review. *Trends in Food Science & Technology* 2016; 49: 96–109.
- **4.** Belwal T., Ezzat S. M., Rastrelli L. et al.: A critical analysis of extraction techniques used for botanicals: trends, priorities, industrial uses and optimization strategies. *Trends in Analytical Chemistry* 2018; 100: 82–102.
- **5.** Picot-Allain C., Mahomoodally M. F, Ak G., and Zengin G.: Conventional versus green extraction techniques—a comparative perspective. *Current Opinion in Food Science* 2021; 40: 144–156.
- 6. Mansinhos I., Goncalves S., Rodriguez-Solana R., Ordonez-Diaz J. L., Moreno-Rojas J. M, and Romano A: Ultrasonic-assisted extraction and natural deep eutectic solvents combination: a green strategy to improve the recovery of phenolic compounds from *Lavandula pedunculata subsp. lusitanica* (chaytor) franco. *Antioxidants* 2021;10(4):582-590.
- 7. Kumar K., Srivastav S., and Sharanagat V. S.: Ultrasound assisted extraction (UAE) of bioactive compounds from fruit and vegeTable processing by-products: a review. *Ultrasonics Sonochemistry* 2021; 70:1-8.
- **8.** Muckensturm B., Foechterlen D., Reduron J. P., Danton P., and Hildenbrand M.: Phytochemical and chemotaxonomic studies of *Foeniculum vulgare*. *Biochemical Systematics and Ecology*1997; 25(4):353–358.
- **9.** Manonmani R. and Abdul Khadir V. M.: Antibacterial screening on *Foeniculum vulgare* Mill. *International Journal of Pharma and Bio Sciences* 2011; 2(4):390–394.
- **10.** Orhan I. E., "Ozc elik B., Kartal M., and Kan Y.: Antimicrobial and antiviral effects of essential oils from selected Umbelliferae and Labiatae plants and individual essential oil components. *Turkish Journal of Biology* 2012;36(3): 239–246.
- **11.** Dua A., Garg G., and Mahajan R.: Polyphenols, flavonoids and antimicrobial properties of methanolic extract of fennel (*Foeniculum vulgare Miller*). European Journal of Experimental Biology 2013; 3(4): 203–208.
- **12.** Ozbek H., U gras, S., D ulger H. et al.: Hepatoprotective effect of *Foeniculum vulgare* essential oil," *Fitoterapia* 2003;74(3): 317–319.
- **13.** Oktay M., G'ulc in I., and "K'ufrevioglu O. I.: Determination of in vitro antioxidant activity of fennel (*Foeniculum vulgare*) seed extracts. *LWT-Food Science and Technology* 2003; 36 (2):263–271.
- **14.** Pradhan M., Sribhuwaneswari S., Karthikeyan D. et al.: In-vitro cytoprotection activity of *Foeniculum vulgare* and *Helicteres isora* in

- cultured human blood lymphocytes and antitumour activity against B16F10 melanoma cell line. *Research Journal of Pharmacy and Technology* 2008;1(4):450–452.
- **15.** El-Soud N. A., El-Laithy N., El-Saeed G. et al.: Antidiabetic activities of *Foeniculum vulgare* mill. Essential oil in streptozotocin-induced diabetic rats. *Macedonian Journal of Medical Sciences* 2011; 4(2):139–146.
- **16.** Koppula S. and Kumar H.: Foeniculum vulgare Mill (Umbelliferae) attenuates stress and improves memory in wister rats. Tropical Journal of Pharmaceutical Research 2013; 12(4):553–558.
- **17.** Shamkant B., Vainav V., and Atmaram H.: *Foeniculum vulgare* Mill: A review of its botany, phytochemistry, pharmacology, contemporary application, and toxicology. *BioMed Research International* 2014;2014(1):1-32.
- **18.** Abdul-Jalil Th. Z.: Screening of rutin from seeds and leaves extract of dill, coriander and fennel cultivated in Iraq. *Pharmacy Global IJCP* 2013; 3(2): 1-16.
- **19.** Torres M., Talavera T.A., Andrews H.E., Contreras A.S. and Pacheco N.: Ultrasound assisted extraction for the recovery of phenolic compounds from vegeTable sources. *Agronomy* 2017; 7(47): 1-19.
- **20.** Abubakar A.R. and Haque M.: Preparation of medicinal plants: basic extraction and fractionation procedures for experimental purposes. *Journal of Pharmacy and Bioallied Sciences* 2020;12(1):1-10.
- 21. Bargah R.K.: Preliminary test of phytochemical screening of crude ethanolic and aqueous extract of *Moringa pterygosperma* Gaertu. *Journal of Pharmacognosy and phytochemistry* 2015; 4(1): 7-9.
- **22.** Hind M. Jewely H.M. and Abdul-Jalil Th.Z.: Extraction, isolation and identification of caffeic Acid and p-coumaric acid from n-butanol fraction of Iraqi *Osteospermum ecklonis* (F. Asteraceae). *IJDDT* 2022; 12 (2),648-653.
- **23.** Hamad M.N.: Detection and isolation of Flavonoids from Calendula officinalis (F. Asteraseae) cultivated in Iraq. *Iraq J Pharm Sci* 2016; 25(2): 1-6.
- **24.** Moulazadeh A., Ranjbar R., Ardestani A.M., Ranjbar K., Akbar Farjadfar A. et al: Cytotoxic effects of *Trachyspermum ammi* and *Ferula assafoetida* on MCF-7 and MDA-MB-468 breast cancer cell lines. *Beni-Suef Univ J Basic Appl Sci* 2022; 11(147):1-10.
- **25.** Ibrahem N. M., Mutlag Sh. H., and Kadhim E.J.: The cytotoxic effect of Iraqi *Rhus Coriaria* against breast and esophagus cancer cells.

- Drugs and Cell Therapies in Haematology 2021;10(3):115-129.
- **26.** Ali Z, Jabir M, Al-Shammari A.: Gold nanoparticles inhibiting proliferation of human breast cancer cell line. *Research journal of biotechnology* 2019; 14:79-82.
- 27. Ogbole OO, Segun PA, Adeniji AJ.: In vitro cytotoxic activity of medicinal plants from Nigeria ethno-medicine on Rhabdomyosarcoma cancer cell line and HPLC analysis of active extracts. *BMC complementary and alternative medicine* 2017:17(1):1-10.
- **28.** Singh Sh. P.: A Comprehensive Review on Pharmacological Activity of Foeniculum vulgare. Global Journal of Pharmacy & Pharmaceutical Sciences 2019;7(1):1-5.
- **29.** Sharopov F., Valiev A., Satyal P., Gulmurodov I., Yusufi S., et al: Cytotoxicity of the essential oil of fennel (*Foeniculum vulgare*) from Tajikistan. *Foods* 2017; 6(73):1-11.
- **30.** AlTemimi A., Lashassassi N., Baharlouei A., Watson D.J. and Lightfoot D.A.: Phytochemicals: extraction, isolation and identification of bioactive compounds from plant extracts. *Plants* 2017; 6(42): 1-23.
- **31.** Xu Y. and Pan S.: Effects of various factors of ultrasonic treatment on the extraction yield of all trans-lycopene from red grape fruit (Citrus paradise Macf.). *Ultrason. Sonochem* 2013; 20: 1026-1032.
- **32.** Kumara K, Srivastava S, Sharanagatb V.S.: Ultrasound assisted extraction (UAE) of bioactive compounds from fruit and vegeTable

- processing by-products: A review. *Ultrason. Sonochem* 2021; 70:1-11.
- **33.** Xu D., Zhou Y., Zhang J., Li S., Li A. and Li H.: Optimization of ultrasound assisted extraction of natural antioxidant from the flower of *Jatropha integerrima* by response service methodology. *Molecules* 2016; 21: 1-18.
- **34.** Yue T., Shao D., Yuan Y., Wang Z. and Qiang C.: Ultrasound assisted extraction, HPLC analysis and antioxidant activity of polyphenols from unripe apple. *J Sep Sci* 2012; 35: 2138-2145.
- **35.** LinaresG and RojasM.L.: Ultrasound-assisted extraction of natural pigments from food processing by-products: A review. *Frontiers in Nutrition* 2022; 9:1-17.
- **36.** Çevik D., Kan Y., and Kırmızıbekmez H.: Mechanisms of action of cytotoxic phenolic compounds from *Glycyrrhiza iconica* roots. *Phytomedicine* 2019; 58:1-12.
- **37.** Rezaei-Seresht H., Cheshomi H., Falanji F., MovahediMotlagh F., Maryam Hashemian M. and Mireskandari E.: Cytotoxic activity of caffeic acid and gallic acid against MCF-7 human breast cancer cells: An *in silico* and *in vitro* study. *AJP* 2019; 9(6):574-586.
- **38.** Zaremba-Czogalla M., Jaromin A., Sidoryk K., Zagórska A., Cybulski M. and Gubernator J.: Evaluation of the *in Vitro* cytotoxic activity of caffeic acid derivatives and liposomal formulation against pancreatic cancer cell lines. *Materials* 2020;13(5813): 1-19.



This work is licensed under a <u>Creative Commons Attribution 4.0 International License</u>.